

Fundación Miguel Lillo Tucumán Argentina



# Mycorrhizal biofertilizer: advantages and hindrances in its application

Biofertilizante micorrízico: ventajas y desventajas en su aplicación

Ghosh, Somdatta<sup>®</sup>; Som S. Dutta<sup>®</sup>; Suvashree Bhaumik<sup>®</sup>; Kunwar Hansda<sup>®</sup>; Debashis Kuila<sup>®</sup>

Mycorrhiza & Ecology Research Section, UG & PG Department of Botany, Midnapore College (Autonomous), Midnapore, West Bengal, 721101, India.

\* Corresponding author: <somsubhradutta92@gmail.com>

### Abstract

Global climate change, rising population growth, and the overuse of agrochemicals have led to various problems, including soil degradation, a decline in food production, and environmental issues affecting the agricultural sector. To overcome these challenges, biofertilizers (particularly, those of mycorrhizal origin) have emerged as a sustainable and eco-friendly alternative. Arbuscular mycorrhiza (AM) is an association between plant roots and fungi from the subphylum Glomeromycotina, found in approximately 72% of land plants, and is one of the most common and ancient types of symbiosis on Earth. AM fungi provide with numerous benefits in exchange for plant photoassimilates. Although AM fungi may represent a viable biofertilizer option, their use is significantly limited due to various issues, ranging from their production to field application. This review sheds light on the potential of AM fungi as biofertilizers, analyzing their diverse benefits while also addressing the limitations associated with their production and application for enhancing agricultural productivity.

**Keywords:** Agricultural productivity; agrochemicals; biofertilizers; global climate change; mycorrhiza.

<sup>➤</sup> Esta obra está bajo una Licencia Creative Commons Atribución – No Comercial – Sin Obra Derivada 4.0 Internacional.





<sup>➤</sup> Ref. bibliográfica: Ghosh, S.; Dutta, S. S.; Bhaumik, S.; Hansda, K.; Kuila, D. 2025. Mycorrhizal biofertilizer: advantages and disadvantages in its application. *Lilloa 62* – Suplemento N° 2: "Interacciones biológicas en un mundo cambiante": 193-221. doi: https://doi.org/10.30550/j.lil/2141

<sup>&</sup>gt; Recibido: 12 de marzo 2025 - Aceptado: 31 de mayo 2025 - Publicado: 27 de junio 2025.

<sup>➤</sup> URL de la revista: http://lilloa.lillo.org.ar

#### Resumen

El cambio climático global, el aumento de la población y el uso excesivo de productos agroquímicos han dado lugar a diversos problemas, como la degradación del suelo, la caída de la producción de alimentos y cuestiones medioambientales que han arrinconado al sector agrícola. Para superarlos, los biofertilizantes, en particular los micorrícicos, han surgido como una alternativa sostenible y respetuosa con el medio ambiente. La micorriza arbuscular (MA), una alianza entre las raíces de las plantas y los hongos del subfilo Glomeromycotina, que se encuentra en aproximadamente el 72% de las plantas terrestres, es uno de los tipos de simbiosis más comunes y antiguos de la Tierra. Los hongos MA proporcionan a las plantas multitud de beneficios a cambio de fotoasimilados vegetales. Aunque los hongos MA pueden ser una buena opción biofertilizante, su uso es muy limitado, ya que existen diversos problemas que van desde su producción hasta su aplicación. Esta revisión arroja luz sobre el potencial de los hongos MA como biofertilizantes, analizando sus diversos beneficios y abordando al mismo tiempo las limitaciones asociadas a su producción y aplicación para aumentar la productividad agrícola.

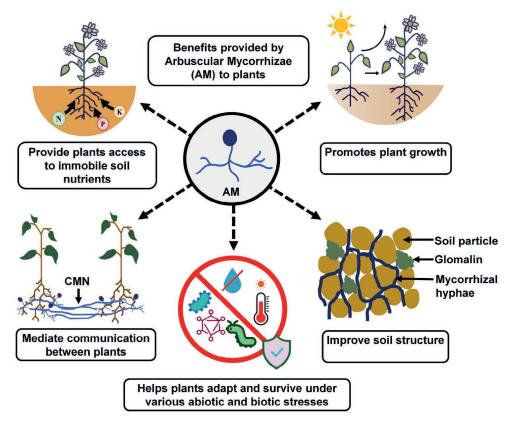
**Palabras clave:** Productividad agrícola; agroquímicos; biofertilizantes; cambio climático global; micorrizas.

#### INTRODUCTION

Global climate change, the exponential rise in the human population, the energy crisis, and the limited availability of natural resources have placed immense pressure on the agricultural sector (Kinge et al., 2022). In addition, the long-term injudicious use of synthetic chemical fertilizers and pesticides to raise agricultural productivity has given rise to many problems, such as deterioration of soil quality, which has resulted in a decrease in food production (Naik et al., 2020; Fasusi et al., 2023) and has given rise to many environmental problems, such as eutrophication of water bodies (Bakhshandeh et al., 2017; Ammar et al., 2023). This, in turn, has led to the degradation of ecosystems and has jeopardized trophic interactions (Barros-Rodríguez et al., 2024). This unprecedented situation has put various stakeholders, such as governments, policymakers, scientists, and farmers, in an alarming state (Stephenson et al., 2013). The growing awareness of the environmental and ecological consequences of heavy reliance on synthetic fertilizers has prompted a shift towards more sustainable agricultural practices to ensure food security for this expanding population while simultaneously conserving Earth's natural resources, with biofertilizers emerging as a viable, eco-friendly, sustainable, and cost-saving solution (Hunter et al., 2017; Nosheen et al., 2021).

Biofertilizers are formulations containing microorganisms or microbial-derived molecules that enhance plant growth and productivity by approximately 10-40% (Shahwar et al., 2023; Ferreyra-Suarez et al., 2024). Among biofertilizers, mycorrhizal biofertilizers have garnered significant attention because of their ability to enhance plant nutrient acquisition and overall growth performance (Kour et al., 2020; Ammar et al., 2023). Mycorrhizae, first reported by Frank (1885), is the mutualistic association between fungal hyphae and plant roots in which there is a reciprocal exchange of nutrients in the form of photo-assimilates like hexose sugars and lipids from plants to fungi and mineral nutrients from fungi to plants (Kaiser et al., 2014; Luginbuehl et al., 2017). Mycorrhizal association can be categorized into four types: arbuscular mycorrhiza (AM), ectomycorrhiza (EcM), orchidioid mycorrhiza (OrM), and ericoid mycorrhiza (ErM) (Brundrett & Tedersoo, 2018). Among these, arbuscular mycorrhiza, a symbiotic association between plant roots and obligate biotrophic fungi of the subphylum Glomeromycotina (Spatafora et al., 2016), is one of the most ancient and widespread symbioses on Earth (Kenrick & Strullu-Derrien, 2014). It is found in approximately 72% of terrestrial land plants (Genre et al., 2020), ranging from bryophytes to angiosperms from a wide range of ecosystems (Redecker et al., 2013), which played a pivotal role in allowing plants to transition from an aquatic environment before the evolution of true roots (Kenrick & Strullu-Derrien, 2014; Kuyper & Jansa, 2023). In contrast, ectomycorrhizal associations are found in only 2% of land plants, primarily those associated with temperate trees (Brundrett & Tedersoo, 2018). The other two types of mycorrhizal associations, Orchidioid and Ericoid, were restricted to specific plant families, Orchidaceae and Ericaceae, respectively (Brundrett & Tedersoo, 2018).

In addition to their widespread occurrence and distribution across ecosystems, arbuscular mycorrhizae provide plants with an array of benefits, such as providing them access to immobile soil nutrients (Smith & Smith, 2011; Yu et al., 2022), promoting plant growth (Nadeem et al., 2014), and helping plants adapt and survive under various abiotic (Begum et al., 2019) and biotic stresses (Dey & Ghosh, 2022). They also improve soil structure through soil aggregation (Gosling et al., 2010) and mediate communication between plants through a Common Mycorrhizal Network (CMN) (Walder et al., 2012; Heklau et al., 2021) (Fig. 1). In addition, they provide various ecosystem services, such as the breakdown of organic matter (Powell & Rillig, 2018), maintenance of belowground microbial diversity, and regulation of plant community diversity (Van Der Heijden et al., 2015; Fall et al., 2022). Thus, they have huge potential to be harnessed as a sustainable biofertilizer option for boosting agricultural productivity, with the potential to reduce reliance on synthetic fertilizers and pesticides while improving plant growth and resilience. However, they remain underutilized owing to inherent challenges associated with mass production, the risk of contamination, the need for skilled labour (Gianinazzi & Vosátka, 2004; Madawala,



**Fig. 1.** Benefits provided by AM (Arbuscular Mycorrhizal) fungi to plants. Common Mycorrhizal Network (CMN).

**Fig. 1.** Beneficios que aportan los hongos MA (micorrizas arbusculares) a las plantas. Red común de micorrizas (CMN).

2021), and conventional agricultural practices, such as the application of agrochemicals, tillage, and crop rotation, which disrupt mycorrhiza development (Brito *et al.*, 2012; Bakhshandeh *et al.*, 2017). Among mycorrhizas, arbuscular mycorrhiza (AM) will be considered in this review, as they are found inclusively in all habitats and form symbiosis with nearly 72% of terrestrial land plants, whereas ectomycorrhizae associate with only 2% of land plants, mostly with trees found in forests of temperate regions (Brundrett & Tedersoo, 2018).

This review elucidates the potential of arbuscular mycorrhizal (AM) fungi as biofertilizers by discussing the numerous benefits they offer. Furthermore, this paper highlights the various challenges that may arise in the application of AM fungi as biofertilizers intended to enhance agricultural productivity.

#### **ADVANTAGES**

The symbiotic association between plants and mycorrhizae provides the plants with an array of benefits and services that promote their growth and, at the same time, help them to cope with various stresses, which are discussed below.

### Facilitation of nutrient uptake

Mycorrhizal colonization induces changes in the morphological characteristics of plant roots that are crucial for hosting mycorrhizal structures within the cells and enhancing nutrient uptake and accumulation by mycorrhizal plants (Neumann & George, 2010). The morphological modifications in the plant root system triggered by mycorrhizal colonization include an increase in total root length, a change in root-shoot ratio, an increase in root branching, and an increase in the number of root tertiary branches (Vierheilig et al., 2008). Along with these root modifications, the hyphae of mycorrhizal fungi have the capacity to expand beyond the root surface by a distance greater than 10 cm (Jakobsen et al., 1992). These hyphae are very thin, with an average diameter of 3 to 4  $\mu$ m, which is much smaller than the finest root hairs, which have an average diameter of  $10 \,\mu\mathrm{m}$  (Johansen et al., 1993); thus, they are able to extend their growth into the tiniest micropores of soil, which enhances their nutrient absorption efficiency (Bennett & Groten, 2022). Altogether, the root modifications induced by AM fungi and the AM hyphae form an extensive nutrient-absorbing network that expands the root zone absorption area by 10% to 100% (Etesami et al., 2021) that stretches beyond the nutrient depletion zones that form around roots in the rhizosphere (The narrow zone of soil surrounding plant roots), which enables the AM-colonized roots to access a larger volume of the soil than roots that are not colonized by AM fungi (Cavagnaro et al., 2015). AM also promotes the expression of phosphate transporters (PTs) in various host plants, including StPT3 in potato (Rausch et al., 2001), MtPT4 in Medicago truncatula (Harrison et al., 2002), and OsPT11 in rice (Paszkowski et al., 2002). These transporters play a critical role in facilitating the uptake of phosphate released by AM fungi at the symbiotic interface into plant cells (Wang et al., 2017). In addition to phosphate transporters, plant ammonium transporters such as GmAMT4.1 and ATM2;3 in soybean and Medicago are induced by the presence of the fungus within arbuscule-containing cortical cells in the roots (Wang et al., 2017).

Mycorrhizal fungi secrete enzymes, like acid phosphatases and proteases, which facilitate the solubilization of both organic and inorganic phosphorus compounds in the soil (Miransari *et al.*, 2009). This enzymatic activity enhances the bioavailability of phosphorus, thereby improving the availability of phosphorus to the plant host (Samantaray *et al.*, 2024).

Some mycorrhizal fungi produce siderophores (Low molecular weight organic compounds that have a high affinity for ferric ions) that complex iron and oxalate to increase potassium absorption from the soil, resulting in improved plant nutrition (Lindahl *et al.*, 2007).

# Improvement of water absorption capacity

Mycorrhizal hyphae are very thin, usually having a diameter of 2–5  $\mu$ m, which is approximately two times smaller than plant roots, which have an average diameter of 10–20  $\mu$ m, which allows them to transport water through small soil cavities inaccessible to plant roots (Diagne *et al.*, 2020). In addition, the hyphal tips are also hydrophilic, which enhances the transport of water from the soil to the plant cells (Miransari, 2011).

AM fungi also produces glomalin, which is a glycoprotein containing 30–40% carbon compounds that is deposited on the soil particles and holds the soil particles together, forming stable soil aggregations that improve the water holding capacity of soil and protect the soil from desiccation (Verbruggen *et al.*, 2012; Sharma *et al.*, 2017).

Mycorrhizal fungi also induce the expression of plant aquaporin genes in mycorrhizal roots, which encode integral membrane transporters that transport water, signalling molecules, and ions through cell membranes (Santander *et al.*, 2021). Aquaporins enhance the water absorption capacity of plants and improve their tolerance to drought stress. Thus, AM fungi improves plants resilience to drought stress through multilayered, controlled protection mechanisms.

# Enhancement of growth and yield of crops

Mycorrhizal symbiosis plays a significant role in enhancing the productivity and quality of tropical agricultural crops, particularly in regions where phosphorus deficiency is prevalent in the soil (Hildermann et al., 2010). Nziguheba and Smolders (2008) stated that 75% of the phosphorus applied to crops is not utilized by plants. Zea mays and flax are very dependent on AM fungi to meet their primary phosphorus requirements (Bai et al., 2003; Thompson et al., 2013). AM fungi inoculation can provide up to 90% of plant phosphorus and 20% of plant nitrogen due to the hyphal network in the soil formed after symbiotic associations with the host plant (Johnson et al., 2012). Most of the major agricultural crops are mycorrhizal hosts and increase the inoculum potential of the soil and colonization of future crops (Schliemann et al., 2008). AM fungi have a widespread distribution, and their use in agroecosystems as mycorrhizae-based inoculants is increasing (Igiehon & Babalola, 2017). In the case of potatoes, AM inoculation was observed to increase the total crop yield by 9.5%.

When AM fungi inoculant (*Rhizophagus irregularis*) was applied to potatoes over a period of four years in North America and Europe under real field conditions and showed a highly significant increase (42.2 tons/ha) in potato yield compared to non-inoculated controls (38.3 tons/ha) (Hijri, 2016). AM fungi also contributes to enhancing crop yield in rainfed agricultural systems by promoting drought resistance in host plants, which is particularly important in mitigating yield losses attributed to phytopathogens and herbivores (Dowarah *et al.*, 2022).

Furthermore, AM fungi have been engaged in large-scale field production of maize (Sabia et al., 2015). The inoculation of the AM fungus Rhizophagus irregularis with the cotton cultivar Lumian No. 1 reduced the requirement of fertilizer application in the field (Gao et al., 2020). Thus, AM fungi possess a considerable potential for enhancing the yield of crops.

A positive relationship between mycorrhizal spore population and fruit yield (number and weight of fruits) has been observed in various fruit trees (Bona et al., 2017). Zeng et al. (2014) observed increased levels of sugars, organic acids, vitamin C, flavonoids, and minerals in citrus fruits due to Glomus versiforme, resulting in improved quality. The inoculation of Glomus macrocarpum, G. coledonicum, and Acaulospora sp. resulted in enhanced plant height, stem diameter, and biomass in trifoliate and troyer oranges (De Souza, 2000). Inoculation with Gigaspora rosea and Glomus mosseae enhanced the growth of different grape rootstocks and cultivars compared to uninoculated plants (Linderman & Davis, 2001). Arbuscular mycorrhizal fungi also increased the yield and productivity of apple trees when plants were co-inoculated with phosphate-solubilizing bacteria (Aslantaş et al., 2007).

# Contributes to soil sustainability

Arbuscular mycorrhizae are an essential component of the pedosphere that regulate important soil processes and are considered to have immense potential for improving soil sustainability (Powell & Rillig, 2018; Zhang *et al.*, 2024). AM fungi contribute to soil sustainability by regulating three major factors: the structure of the soil, physiological processes in plants, and ecological dynamics (Fall *et al.*, 2022).

Mycorrhizal fungi form a large amount of mycelia in the soil, which continuously regenerates and forms a matrix that wraps and interconnects soil particles, improving the stability of soil aggregates, minimizing soil compaction, and improving the water-holding capacity of soil (Chen et al., 2018). Mycorrhizal fungi also secrete a negatively charged, hydrophobic, and thermotolerant glycoprotein, glomalin (Fall et al., 2022). Glomalin acts as a glue that binds soil particles and stabilizes soil aggregation (Lehmann et al., 2020). Furthermore, the hydrophobic nature of glomalin provides soil aggregates with water resistance. Additionally, glomalin is slowly biodegradable by soil microorganisms (Hu et al., 2019).

Together, the formation of soil aggregates by the enmeshment action of hyphae and the increase in their stability by glomalin minimize the risk of soil compaction and increase the water-holding capacity of the soil, which results in reduced soil erosion, nutrient leaching, and denitrification, thereby improving soil fertility (Pellegrino *et al.*, 2020). The mycelial network, in addition to forming stable soil aggregates, also contributes to the formation of soil organic matter after death (Hawkins *et al.*, 2023).

Thus, AM fungi participate in various types of essential soil functions, such as nutrient cycling (Powell & Rillig, 2018; Frey, 2019), reducing soil nutrient loss by minimizing nutrient leaching from the soil (Cavagnaro et al., 2015), and improving the soil structure by producing a hydrophobic glycoprotein glomalin (Leifheit et al., 2014). AM also regulates various physiological processes in plants, such as enhancing their nutrient acquisition capacity from the soil (Rouphael et al., 2015), modulating phytohormone levels in plants, and reprogramming the secondary metabolism of plants (Rivero et al., 2015). AM also influences the ecological dynamics of soil by recruiting beneficial soil microorganisms, such as phosphate-solubilizing bacteria (PSB), nitrogen-fixing bacteria, and plant growth-promoting rhizobacteria (PGPR), to the mycorrhizosphere (The zone of soil surrounding the plant roots colonized by mycorrhizal fungi) (Yu et al., 2022).

# Strengthens plant immunity to biotic stress

AM induces disease control by both indirect and direct means. Indirectly through nutrition improvement; synthesis of plant hormones (Song *et al.*, 2015) and competing with other harmful microbes on the root surface and within the root. AM fungi also produce some antifungal (Bencherif *et al.*, 2019) and antibacterial compounds (Kaur & Sussella, 2020) and toxins that act against pathogenic organisms (Wang *et al.*, 2018).

AM fungal symbiosis directly inhibits pathogens by mycorrhiza-induced resistance (MIR) (Nguvo & Gao, 2019) by creating systemic protection against a wide range of pathogens. MIR includes characteristics of both systemic acquired resistance (SAR), which occurs after pathogen infection in plants, and induced systemic resistance (ISR), which occurs following root colonization by non-pathogenic rhizobacteria (Cameron *et al.*, 2013). MIR activates both pathogen-specific and broad-range defence genes (Fiorilli *et al.*, 2018) to produce enzymes and pathogenesis-related (PR) proteins (Sanmartín *et al.*, 2020).

AM fungi are active against different types of nematodes (Da Silva Campos, 2020), bacteria (Sanmartín *et al.*, 2020), viruses (Aseel *et al.*, 2019), and fungi (Song *et al.*, 2015) in different hosts, though the protective effects vary with AM and host species or other conditions.

#### Enhancement of abiotic stress tolerance

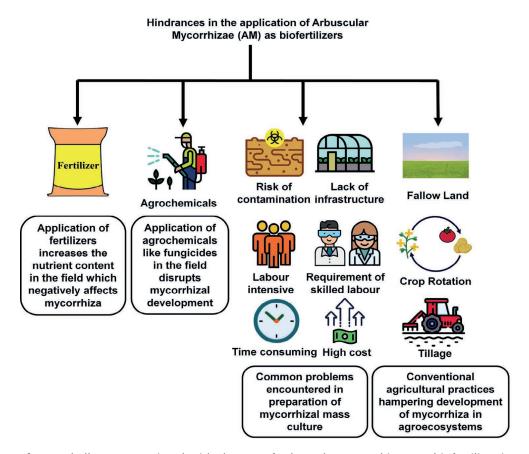
Water stress, caused by drought and salinity, is one of the main abiotic stresses that impact plant growth and productivity. The symbiosis between plant roots and AM fungi is a common strategy for adapting to water stress (Brachmann & Parniske, 2006). Mycorrhizal fungi employ various strategies to sustain host vitality during water stress. Water stress decreases the turgor pressure and water potential of plant cells, which results in the formation of reactive oxygen species like superoxide and hydroxyl radicals (Laxa et al., 2019). As mycorrhizal hyphae can explore a large volume of the soil, they enhance plants water absorption, which improves turgor potential, stomatal conductance, and hence elevates transpiration rate (Augé et al., 2015). AM colonization also improves osmotic balance by inducing accumulation of osmo-protectants such as proline, polyamines, glycine betaine, non-structural carbohydrates, and inorganic solutes like K<sup>+</sup>, Ca<sup>2+</sup>, and Mg<sup>2+</sup> (Baslam & Goicoechea, 2012; Yooyongwech et al., 2013) inside plant cells, which maintains the turgor pressure and protects plant cells from the adverse effects of water stress (Grümberg et al., 2015).

Under salinity stress conditions, AM fungi enhance the uptake of nutrients like phosphorus (P), nitrogen (N), potassium (K), zinc (Zn), and copper (Cu) and maintain ionic homeostasis (Hanin et al., 2016). AM fungi also enhance the levels of the abiotic stress hormone abscisic acid (ABA) in plants to cope with the detrimental effects of water and salinity stress (Martín-Rodríguez et al., 2016). ABA regulates transpiration rates, stomatal movements, root hydraulic conductivity, and the expression of aquaporin-encoding genes (Ouledali et al., 2019). ABA triggers stomatal closure, which in turn reduces transpiration and minimizes water loss during drought stress (Chitarra et al., 2016). AM fungi also augment antioxidant activities to defend against damage by reactive oxygen species (ROS) and promote photosynthesis to minimize the detrimental effects of salts on the growth and development of plants (Evelin et al., 2009). Crop plants inoculated with AM fungi have been reported to enhance growth and yield, while mitigating osmotic and ionic imbalances to normal levels, allowing crops to thrive under salinity stress (Hanin et al., 2016).

All these adjustments by AM fungi improve plant resilience to various abiotic and biotic stresses, which improves plant growth and ultimately their productivity. Thus, AM can be utilized as a bioinoculant in the soil, which can maximize the output without compromising soil health while simultaneously ensuring soil sustainability.

#### HINDRANCES IN APPLICATION

Despite the multitude of benefits offered by mycorrhiza, their widespread adoption and successful implementation as biofertilizers in modern agriculture face several challenges (Fig. 2).



**Fig. 2.** Challenges associated with the use of arbuscular mycorrhiza as a biofertilizer in agroecosystems.

Fig. 2. Desafíos asociados al uso de micorrizas arbusculares como biofertilizante en agroecosistemas.

# High nutrient content in soil

Under nutrient-poor conditions in the soil, plant roots release strigolactones (A family of carotenoid-derived phytohormones that promote seed germination in parasitic plants and facilitate the establishment of symbiosis between plants and AM fungi) into the soil, which induces the germination of AM spores and stimulate hyphal branching (Waters et al., 2017). The AM fungi in response release myc factors that are perceived by plant receptors, resulting in the induction of the common symbiotic pathway leading to the establishment of symbiosis (Zhang et al., 2015). However, in agroecosystems with high fertilizer input, the level of nutrients, especially phosphorus (P), becomes high, and plants can take up phosphorus (P) from the soil without seeking any help from the AM fungi, and the symbiotic association transforms to a parasitic one in contrast to mutualism under low soil N and P levels (López-Ráez et al., 2017). Therefore, it is no longer feasible for plants to remain in this association, as there is a carbon cost; thus, the allocation of carbohydrates and lipids to AM fungi is reduced (Qin et al., 2024).

Additionally, the release of strigolactones from plant roots decreases (López-Ráez et al., 2011). Finally, plants also downregulates the expression of phosphate transporter (PT) genes (Sawers et al., 2017). As a result, the AM fungal colonization and sporulation are reduced (Tiamtanong et al., 2015). In addition to phosphorus (P), the mycorrhizal community is greatly affected by chronic nitrogen deposition. Many nitrogen fertilizers have been reported to decrease colonization in both field and pot experiments (Getman-Pickering et al., 2021). Nitrogen fertilization alone or in combination with phosphorus disrupts AM symbiotic efficiency. Potassium (K) more than the natural soil K content decreased mycorrhizal colonization in maize (Ardestani et al., 2011).

# Application of agrochemicals

Agrochemicals (fungicides, pesticides etc.) are now integral part of technology dependent modern conventional agriculture as most high yielding crops are more susceptible to diseases than their wild genotypes. Systemic fungicides have selectivity and specificity for certain pathogens while non-systemic fungicides are broad spectrum and kill all organism exposed to these. Common non-systemic fungicides like *pentachloronitrobenzene* (PCNB), *thiram*, *fotran*, *arsan*, *langstan*, *Chlorothalinol*, *Captapol*, *chloroneb*, *metaxyl* and *ethazole* are highly toxic; while Captan, *Mylone*, *Vapram* and *Volax* are moderately toxic to mycorrhizal fungi. Daconil, Sodium azide, terrazole, captain, and copper sulphate may favour AM activity and development at low doses under specific environmental conditions (Vyas & Vyas, 2000).

Systemic fungicides are more detrimental to AM fungi as they get accumulated inside the roots (Jin et al., 2013). As systemic fungicides are mostly fungi static, they have less effect on spore germination and hyphal growth, but they affect infection, colonization and sporulation. Benomyl, Tridemorph, Triforine, Ethirimol, Etridiazole, Thiophanate methyl, Thiabendazole, Thiademifom and Carboxin show detrimental effects on the development of AM in the root (Wang et al., 2018). Almost all non-systematic fungicides adversely affect AM and are retained in soil (Ghosh et al., 2024).

# Selection and preparation of mass culture

The primary challenge in producing an AM fungi inoculum is the obligate symbiotic nature of AM fungi, which requires a host plant to grow and complete its life cycle. Thus, they cannot be cultivated in pure culture without their host plants (Säle et al., 2015). Consequently, their propagation must include a cultivation phase with host plants maintained in fields, greenhouses, or growth chambers. This is labour intensive, costly, and at the same time requires considerable time and space (Gianinazzi &

Vosátka, 2004). Additionally, AM fungal inoculum is a combination of soil and AM propagules; therefore, these inocula cannot be entirely free from contamination with pathogens and weedy seeds (Kumar et al., 2017). Furthermore, the soil makes AM inocula bulkier, and their transport becomes more challenging and expensive (Ceballos et al., 2013). Apart from these evident limitations associated with the production of AM inocula on a large scale, there are additional obstacles like lack of infrastructure for inoculum production, storage, and skilled labour, as well as lack of a suitable carrier material, short shelf life, and inconsistency in the inoculum (Mukhongo et al., 2016). Additionally, there is a lack of quality control protocols for AM fungal inoculum production; thus, the species listed in commercial products may not be entirely accurate (Hart et al., 2018). Again, not all AM fungi species may be suitable for all hosts, soil types, or climates (Jansa et al., 2014). Thus, screening is necessary.

# Conventional agricultural practices like fallowing land, rotation with non-host plant and tillage

Conventional agricultural practices, aside from the application of agrochemicals, such as tillage and crop rotation, which inhibit the development of mycorrhizae, particularly in topsoil layers, are two commonly encountered challenges that prohibit the effective utilization of mycorrhizae as a biofertilizer in agroecosystems.

As an obligate symbiont, prolonged fallowing of land devoid of vegetation or with non-host vegetation leads to the depletion of arbuscular mycorrhizal (AM) propagules in the soil as AM spores are unable to germinate and proliferate in the absence of a host. Similarly, the cultivation of non-host crops, such as those belonging to the Brassicaceae family, which release glucosinolates into the soil, adversely affects AM propagules. Upon their release, glucosinolates decompose into isothiocyanates, which are antimicrobial compounds that further reduce the prevalence of AM propagules in the soil, even following the rotation with host plants (Kirkegaard & Sarwar, 1998; Ghosh *et al.*, 2004).

Intensive tillage disrupts the hyphal networks of mycorrhizae within the soil (Feilinezhad *et al.*, 2022) which can selectively affect various arbuscular mycorrhizal fungi (AMF) groups based on their life and colonization strategies, either promoting or impairing specific groups, leading to a 40% reduction in AM fungal diversity (Brito *et al.*, 2012). Furthermore, mycorrhizal root colonization consistently demonstrates lower levels under tillage conditions compared to no-tillage environments (Castillo, 2006). During the initial stages of colonization, the direct impacts of conventional tillage systems are attributed to the physical disruption of the extraradical mycelium network, resulting in a slowdown of symbiotic efficiency. This disruption hampers AM activity related to nutrient and water uptake, glo-

malin-associated soil aggregate formation (Brito et al., 2012), and bioprotection against soil pathogens (Patanita et al., 2020).

Thus, conventional agricultural practices, aside from the application of agrochemicals, such as tillage and crop rotation, can inhibit the development of mycorrhizae in different agroecosystems. However, adopting practical alternatives such as utilizing organic fertilizers (manures and compost) and slow-release mineral fertilizers (like rock phosphate) (Cavagnaro, 2014), implementing reduced tillage (Ghorui et al., 2024), and shortening the fallow period through crop rotation with mycorrhizal-dependent cover crops such as *Vicia villosa* Roth. and *Trifolium* spp. instead of non-mycorrhizal hosts like Brassicaceae can stimulate arbuscular mycorrhizal (AM) fungi in the field (Njeru et al., 2015). Furthermore, the application of fungicides that do not negatively affect mycorrhizal fungi can lead to an increase in AM propagules and their species diversity (Rouphael et al., 2015).

# Methods of plant inoculation with mycorrhizal fungi

Inoculating plants with AM fungi employs advanced techniques that can significantly enhance plant health and productivity. These methods can be broadly categorized as follows.

**Seed inoculation.**— This method consists of coating seeds with a slurry of AM spores, ensuring that upon germination, the seeds are immediately exposed to the AM fungi (Kafle *et al.*, 2019).

**Direct root inoculation.**— This approach involves dipping the roots of seedlings into a suspension of AM spores during transplanting, thereby establishing direct contact between the AM fungi and the root system (Eulenstein *et al.*, 2017).

**Soil inoculation.**— This strategy involves broadcasting granular or powdered inoculum over the soil surface at the time of tillage or planting, making it suitable for large-scale agricultural operations where direct root inoculation may not be feasible (Aliyu *et al.*, 2019).

While these methods demonstrate potential for enhancing soil health and agricultural productivity, challenges persist in standardizing these practices.

# Dual application of AM fungi and plant growth promoting bacteria (PGPB)

The simultaneous application of Arbuscular Mycorrhizal (AM) fungi and Plant Growth Promoting Bacteria (PGPB), represents an effective ecological strategy to enhance plant performance and soil health compared to single

inoculation (Feng *et al.*, 2023). AM fungi colonize the roots of terrestrial plants; on the other hand, PGPB colonizes the rhizosphere. The combination of these two groups can lead to improved plant performance by providing numerous benefits, such as nitrogen fixation, phosphate solubilization and mineralization, phytohormone production, and enhanced tolerance to various biotic and abiotic stresses (Wahid *et al.*, 2022).

The mechanism behind this synergistic effect is attributed to several factors, including PGPB enhancing AM spore germination and hyphal growth, which leads to more efficient mycorrhizal colonization (Sagar *et al.*, 2021). In return, the PGPB receive nourishment in the form of carbon-rich exudates from the AM fungi. This mutual enhancement results in a more robust root system that better absorbs nutrients and water, promoting plant growth and improving performance under various stresses (Wahid *et al.*, 2022).

The synergistic effect of dual inoculation with AM fungi and PGPB has been validated by several scientific studies. For example, in two wheat (Triticum aestivum) cultivars, HD-3086 and HD-2967, the co-application of Bacillus subtilis CP4 and the AM fungi Glomus fasciculatum significantly increased plant biomass and yield compared to single inoculation and uninoculated controls (Yadav et al., 2021). Additionally, inoculating a strawberry variety (Fragaria ananassa var. Selva) with a consortium of AM fungi (Rhizophagus intraradices, Glomus aggregatum, G. viscosum, Claroideoglomus etunicatum, and C. claroideum) and Pseudomonas fluorescens Pf4 resulted in earlier flowering and fruiting, as well as increased yield and nutritional content (Bona et al., 2015). In Melissa officinalis L., dual inoculation with the AM fungi Glomus mosseae and the PGPB Azospirillum brasilense Sp245 promoted plant growth and yield under water deficit conditions (Gorgi et al., 2022). El-Sharkawy et al. (2022) found that inoculating Rhizophagus irregularis with the bacterium Streptomyces viridosporus HH1 improved growth parameters and induced defence responses in pea (Pisum sativum L.) against infection of Fusarium oxysporum f.sp. pisi.

#### CONCLUSION

The use of mycorrhizae as biofertilizers is a cost-effective strategy for supplying essential nutrients and water and enhancing resilience to various environmental stresses, thereby contributing to sustainable, eco-friendly production that minimizes the use of agrochemicals and reduces environmental and human health risks. To fully harness the potential of AM fungi in enhancing the productivity of agroecosystems, it is essential to troubleshoot the negative effects of conventional agricultural practices on AM fungi, soil quality, ecosystem functioning, and human health. Therefore, to maximize the benefits of AM fungi, it is essential to develop an integrated management system, which is a comprehensive strategy that

combines multiple agricultural practices to increase the number, variety, and functioning of AM fungi. This enhancement aims to improve their symbiotic interaction of AM fungi with their host plants, thereby optimizing mycorrhizal advantages, leading to improved crop performance and agroecosystem sustainability while remaining within economic boundaries.

To achieve such a sustainable system, soil management practices like no-tillage farming or reduced tillage should be adopted, and cover crops should be implemented to promote soil aggregation and increase organic matter content in fields, creating a more favourable environment for mycorrhizal fungi, which will facilitate their growth and colonization of plant roots within agroecosystems. Besides soil management practices, nutrient management strategies should include the judicious application of fertilizers tailored to the specific needs of the crops and the existing soil nutrient levels to mitigate the suppression of mycorrhizal activity caused by excessive fertilization. Also, selecting crop varieties known to be highly responsive to mycorrhizal colonization is crucial to ensure that plants can successfully capitalize on the benefits provided by AM fungi.

An integrated management system should also incorporate strategies to introduce beneficial mycorrhizal fungi populations in the soil, taking into account the broader ecological context of the agroecosystem. This can be achieved by exploring native AM fungi and plant growth-promoting bacteria instead of relying on commercially available bioinoculants, as native populations are pre-adapted to the environment and can establish themselves better in the soil without promoting weed growth, which is often a concern with commercial bioinoculants (Duell *et al.*, 2022). Furthermore, fostering collaborative partnerships between government, researchers, and farmers is vital to facilitate the exchange of knowledge and best practices related to mycorrhizal management.

Additionally, investment in research and development for enhancing the infrastructure for propagule production and building a skilled workforce in this domain are critical to ensure their widespread adoption and success.

#### **CONFLICT OF INTEREST**

The authors declare no potential conflict of interest.

#### **BIBLIOGRAPHY**

- Aliyu, I. A., Yusuf, A. A., Uyovbisere, E. O., Masso, C. & Sanders, I. R. (2019). Effect of co-application of phosphorus fertilizer and in vitro-produced mycorrhizal fungal inoculants on yield and leaf nutrient concentration of cassava. *PLoS ONE* 14 (6): e0218969. https://doi.org/10.1371/journal.pone.0218969
- Ammar, E. E., Rady, H. A., Khattab, A. M., Amer, M. H., Mohamed, S. A., Elodamy, N. I., Al-Farga, A. & Aioub, A. A. A. (2023). A comprehensive overview of eco-friendly bio-fertilizers extracted from living organisms. *Environmental Science and Pollution Research* 30 (53): 113119-113137. https://doi.org/10.1007/s11356-023-30260-x
- Ardestani, N. K., Zare-Maivan, H. & Ghanati, F. (2011). Effect of different concentrations of potassium and magnesium on mycorrhizal colonization of maize in pot culture. *African Journal of Biotechnology* 10 (73). https://doi.org/10.5897/ajb11.556
- Aseel, D. G., Rashad, Y. M. & Hammad, S. M. (2019). Arbuscular Mycorrhizal Fungi Trigger Transcriptional Expression of Flavonoid and Chlorogenic Acid Biosynthetic Pathways Genes in Tomato against Tomato Mosaic Virus. *Scientific Reports* 9 (1). https://doi.org/10.1038/s41598-019-46281-x
- Aslantaş, R., Çakmakçi, R. & Şahin, F. (2007). Effect of plant growth promoting rhizobacteria on young apple tree growth and fruit yield under orchard conditions. *Scientia Horticulturae* 111 (4): 371-377. https://doi.org/10.1016/j.scienta.2006.12.016
- Augé, R. M., Toler, H. D. & Saxton, A. M. (2015). Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. *Mycorrhiza* 25 (1): 13-24. https://doi.org/10.1007/s00572-014-0585-4
- Bai, Z., Li, H., Yang, X., Zhou, B., Shi, X., Wang, B., Li, D., Shen, J., Chen, Q., Qin, W., Oenema, O. & Zhang, F. (2013). The critical soil P levels for crop yield, soil fertility and environmental safety in different soil types. *Plant Soil* 372: 27-37. https://doi.org/10.1007/s11104-013-1696-y
- Bakhshandeh, S., Corneo, P. E., Mariotte, P., Kertesz, M. A. & Dijkstra, F. A. (2017). Effect of crop rotation on mycorrhizal colonization and wheat yield under different fertilizer treatments. *Agriculture Ecosystems & Environment* 247: 130-136. https://doi.org/10.1016/j.agee.2017.06.027
- Barros-Rodríguez, A., Pacheco, P., Peñas-Corte, M., Fernández-González, A. J., Cobo-Díaz, J. F., Enrique-Cruz, Y. & Manzanera, M. (2024). Comparative Study of *Bacillus*-Based Plant Biofertilizers: A proposed index. *Biology* 13 (9): 668. https://doi.org/10.3390/biology13090668

- Baslam, M. & Goicoechea, N. (2012). Water deficit improved the capacity of arbuscular mycorrhizal fungi (AMF) for inducing the accumulation of antioxidant compounds in lettuce leaves. *Mycorrhiza* 22 (5): 347-359. https://doi.org/10.1007/s00572-011-0408-9
- Begum, N., Qin, C., Ahanger, M. A., Raza, S., Khan, M. I., Ashraf, M., Ahmed, N. & Zhang, L. (2019). Role of arbuscular mycorrhizal fungi in plant Growth Regulation: Implications in abiotic stress Tolerance. *Frontiers in Plant Science* 10. https://doi.org/10.3389/fpls.2019.01068
- Bencherif, K., Djaballah, Z., Brahimi, F., Boutekrabt, A., Dalpè, Y. & Sahraoui, A. L. (2019). Arbuscular mycorrhizal fungi affect total phenolic content and antimicrobial activity of *Tamarix gallica* in natural semi-arid Algerian areas. *South African Journal of Botany* 125: 39-45. https://doi.org/10.1016/j.sajb.2019.06.024
- Bennett, A. E. & Groten, K. (2022). The Costs and benefits of Plant–Arbuscular mycorrhizal fungal interactions. *Annual Review of Plant Biology* 73 (1): 649-672. https://doi.org/10.1146/annurev-arplant-102820-124504
- Bona, E., Lingua, G., Manassero, P., Cantamessa, S., Marsano, F., Todeschini, V., Copetta, A., D'Agostino, G., Massa, N., Avidano, L., Gamalero, E. & Berta, G. (2015). AM fungi and PGP pseudomonads increase flowering, fruit production, and vitamin content in strawberry grown at low nitrogen and phosphorus levels. *Mycorrhiza* 25 (3): 181-193. https://doi.org/10.1007/s00572-014-0599-y
- Bona, E., Cantamessa, S., Massa, N., Manassero, P., Marsano, F., Copetta, A., Lingua, G., D'Agostino, G., Gamalero, E. & Berta, G. (2017). Arbuscular mycorrhizal fungi and plant growth-promoting pseudomonads improve yield, quality and nutritional value of tomato: a field study. *Mycorrhiza* 27 (1): 1-11. https://doi.org/10.1007/s00572-016-0727-y
- Brachmann, A. & Parniske, M. (2006). The most widespread symbiosis on Earth. *PLoS Biology* 4 (7): e239. https://doi.org/10.1371/journal.pbio.0040239
- Brito, I., Goss, M. J., De Carvalho, M., Chatagnier, O. & Van Tuinen, D. (2012). Impact of tillage system on arbuscular mycorrhiza fungal communities in the soil under Mediterranean conditions. *Soil and Tillage Research* 121: 63-67. https://doi.org/10.1016/j.still.2012.01.012
- Brundrett, M. C. & Tedersoo, L. (2018). Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytologist* 220 (4): 1108-1115. https://doi.org/10.1111/nph.14976
- Cameron, D. D., Neal, A. L., Van Wees, S. C. & Ton, J. (2013). Mycorrhiza-induced resistance: more than the sum of its parts?. *Trends in Plant Science* 18 (10): 539-545. https://doi.org/10.1016/j.tplants.2013.06.004
- Castillo, C. G., Rubio, R., Rouanet, J. L. & Borie, F. (2006). Early effects of tillage and crop rotation on arbuscular mycorrhizal fungal propagules in an Ultisol. *Biology and Fertility of Soils* 43 (1): 83-92. https://doi.org/10.1007/s00374-005-0067-0

- Cavagnaro, T. R. (2014). Impacts of compost application on the formation and functioning of arbuscular mycorrhizas. *Soil Biology and Biochemistry* 78: 38-44. https://doi.org/10.1016/j.soilbio.2014.07.007
- Cavagnaro, T. R., Bender, S. F., Asghari, H. R. & Van Der Heijden, M. G. (2015). The role of arbuscular mycorrhizas in reducing soil nutrient loss. *Trends in Plant Science* 20 (5): 283-290. https://doi.org/10.1016/j.tplants.2015.03.004
- Ceballos, I., Ruiz, M., Fernández, C., Peña, R., Rodríguez, A. & Sanders, I. R. (2013). The *In Vitro* Mass-Produced Model Mycorrhizal Fungus, *Rhizophagus irregularis*, Significantly Increases Yields of the Globally Important Food Security Crop Cassava. *PLoS ONE* 8 (8): e70633. https://doi.org/10.1371/journal.pone.0070633
- Chen, M., Arato, M., Borghi, L., Nouri, E. & Reinhardt, D. (2018). Beneficial services of arbuscular mycorrhizal fungi From ecology to Application. Frontiers in Plant Science 9. https://doi.org/10.3389/fpls.2018.01270
- Chitarra, W., Pagliarani, C., Maserti, B., Lumini, E., Siciliano, I., Cascone, P., Schubert, A., Gambino, G., Balestrini, R. & Guerrieri, E. (2016). Insights on the impact of arbuscular mycorrhizal symbiosis on tomato tolerance to water stress. *Plant Physiology* 171 (2): 1009-1023. https://doi.org/10.1104/pp.16.00307
- Da Silva Campos, M. A. (2020). Bioprotection by arbuscular mycorrhizal fungi in plants infected with *Meloidogyne* nematodes: A sustainable alternative. Crop Protection135: 105203. https://doi.org/10.1016/j.cropro.2020.105203
- De Souza, P. V. D. (2000). Effect of arbuscular mycorrhizae and gibberellic acid interactions on vegetative growth of Carrizo citrange seedlings. *Ciência Rural* 30 (5): 783-787. https://doi.org/10.1590/s0103-84782000000500007
- Dey, M. & Ghosh, S. (2022). Arbuscular mycorrhizae in plant immunity and crop pathogen control. *Rhizosphere* 22: 100524. https://doi.org/10.1016/j.rhisph.2022.100524
- Diagne, N., Ngom, M., Djighaly, P. I., Fall, D., Hocher, V. & Svistoonoff, S. (2020). Roles of arbuscular mycorrhizal fungi on plant growth and performance: Importance in biotic and abiotic stressed regulation. *Diversity* 12 (10): 370. https://doi.org/10.3390/d12100370
- Dowarah, B., Gill, S. S. & Agarwala, N. (2022). Arbuscular mycorrhizal fungi in conferring tolerance to biotic stresses in plants. *Journal of Plant Growth Regulation* 41 (4): 1429-1444. https://doi.org/10.1007/s00344-021-10392-5
- Duell, E. B., Cobb, A. B. & Wilson, G. W. T. (2022). Effects of commercial arbuscular mycorrhizal inoculants on plant productivity and Intra-Radical Colonization in Native Grassland: Unintentional De-Coupling of a symbiosis? *Plants* 11 (17): 2276. https://doi.org/10.3390/plants11172276

- El-Sharkawy, H. H. A., Rashad, Y. M. & Elazab, N. T. (2022). Synergism between *Streptomyces viridosporus* HH1 and *Rhizophagus irregularis* Effectively Induces Defense Responses to Fusarium Wilt of Pea and Improves Plant Growth and Yield. *Journal of Fungi* 8 (7): 683. https://doi.org/10.3390/jof8070683
- Etesami, H., Jeong, B. R. & Glick, B. R. (2021). Contribution of arbuscular mycorrhizal fungi, Phosphate–Solubilizing bacteria, and silicon to P uptake by plant. *Frontiers in Plant Science* 12. https://doi.org/10.3389/fpls.2021.699618
- Eulenstein, F., Tauschke, M., Behrendt, A., Monk, J., Schindler, U., Lana, M. & Monk, S. (2017). The application of mycorrhizal fungi and organic fertilisers in horticultural potting soils to improve water use efficiency of crops. *Horticulturae* 3 (1): 8. https://doi.org/10.3390/horticulturae3010008
- Evelin, H., Kapoor, R. & Giri, B. (2009). Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. *Annals of Botany* 104 (7): 1263-1280. https://doi.org/10.1093/aob/mcp251
- Fall, A. F., Nakabonge, G., Ssekandi, J., Founoune-Mboup, H., Apori, S. O., Ndiaye, A., Badji, A. & Ngom, K. (2022). Roles of arbuscular mycorrhizal fungi on soil fertility: contribution in the improvement of physical, chemical, and biological properties of the soil. *Frontiers in Fungal Biology* 3. https://doi.org/10.3389/ffunb.2022.723892
- Fasusi, O. A., Babalola, O. O. & Adejumo, T. O. (2023). Harnessing of plant growth-promoting rhizobacteria and arbuscular mycorrhizal fungi in agroecosystem sustainability. *CABI Agriculture and Bioscience* 4 (1). https://doi.org/10.1186/s43170-023-00168-0
- Feilinezhad, A., Mirzaeiheydari, M., Babaei, F., Maleki, A. & Rostaminya, M. (2022). The effect of tillage, organic matter and mycorrhizal fungi on efficiency and productivity use of nutrients in maize. *Communications in Soil Science and Plant Analysis* 53 (20): 2719-2733. https://doi.org/10.1080/00103624.2022.2072869
- Feng, Z., Liu, X., Qin, Y., Feng, G., Zhou, Y., Zhu, H. & Yao, Q. (2023). Cooperation of arbuscular mycorrhizal fungi and bacteria to facilitate the host plant growth dependent on soil pH. *Frontiers in Microbiology* 14: 1116943. https://doi.org/10.3389/fmicb.2023.1116943
- Ferreyra-Suarez, D., García-Depraect, O. & Castro-Muñoz, R. (2024). A review on fungal-based biopesticides and biofertilizers production. *Ecotoxicology and Environmental Safety* 283: 116945. https://doi.org/10.1016/j.ecoenv.2024.116945
- Fiorilli, V., Vannini, C., Ortolani, F., Garcia-Seco, D., Chiapello, M., Novero, M., Domingo, G., Terzi, V., Morcia, C., Bagnaresi, P., Moulin, L., Bracale, M. & Bonfante, P. (2018). Omics approaches revealed how arbuscular mycorrhizal symbiosis enhances yield and resistance to leaf pathogen in wheat. *Scientific Reports* 8 (1). https://doi.org/10.1038/s41598-018-27622-8

- Frank, A. B. (1885). Ueber die auf Wurzelsymbiose beruhende Ernährung gewisser Bäume durch unterirdische Pilze. Berichte Der Deutschen Botanischen Gesellschaft 3: 128-145.
- Frey, S. D. (2019). Mycorrhizal fungi as mediators of soil organic matter dynamics. *Annual Review of Ecology Evolution and Systematics* 50 (1): 237-259. https://doi.org/10.1146/annurev-ecolsys-110617-062331
- Gao, X., Guo, H., Zhang, Q., Guo, H., Zhang, L., Zhang, C., Gou, Z., Liu, Y., Wei, J., Chen, A., Chu, Z. & Zeng, F. (2020). Arbuscular mycorrhizal fungi (AMF) enhanced the growth, yield, fiber quality and phosphorus regulation in upland cotton (*Gossypium hirsutum L.*). *Scientific Reports* 10 (1). https://doi.org/10.1038/s41598-020-59180-3
- Genre, A., Lanfranco, L., Perotto, S. & Bonfante, P. (2020). Unique and common traits in mycorrhizal symbioses. *Nature Reviews Microbiology* 18 (11): 649-660. https://doi.org/10.1038/s41579-020-0402-3
- Getman-Pickering, Z. L., Stack, G. M. & Thaler, J. S. (2021). Fertilizer quantity and type alter mycorrhizae-conferred growth and resistance to herbivores. *Journal of Applied Ecology* 58 (5): 931-940. https://doi.org/10.1111/1365-2664.13833
- Ghorui, M., Chowdhury, S., Balu, P. & Burla, S. (2024). Arbuscular Mycorrhizal inoculants and its regulatory landscape. *Heliyon* 10 (9): e30359. https://doi.org/10.1016/j.heliyon.2024.e30359
- Ghosh, S., Bhattacharya, D. & Verma, N. K. (2004). Mustard (*Brassica campestris* L.) cultivation reduces the vesicular-arbuscular mycorrhizal advantage of successive crops. *Mycorrhiza News* 16 (2): 12-14.
- Ghosh, S., Bhowmik, S. & Dutta, S. S. (2024). Challenges in Application of Arbuscular Mycorrhizal Inocula in Conventional Agriculture. In: Parihar, M., Rakshit, A., Adholeya, A. and Chen, Y (Eds.), Arbuscular Mycorrhizal Fungi in Sustainable Agriculture: Inoculum Production and Application. Singapore, Springer, (pp. 229–252). https://doi.org/10.1007/978-981-97-0296-1 11
- Gianinazzi, S. & Vosátka, M. (2004). Inoculum of arbuscular mycorrhizal fungi for production systems: science meets business. *Canadian Journal of Botany* 82 (8): 1264-1271. https://doi.org/10.1139/b04-072
- Gorgi, O. E., Fallah, H., Niknejad, Y. & Tari, D. B. (2022). Effect of Plant growth promoting rhizobacteria (PGPR) and mycorrhizal fungi inoculations on essential oil in *Melissa officinalis* L. under drought stress. *Biologia* 77 (1): 11-20. https://doi.org/10.1007/s11756-021-00919-2
- Gosling, P., Ozaki, A., Jones, J., Turner, M., Rayns, F. & Bending, G. D. (2010). Organic management of tilled agricultural soils results in a rapid increase in colonisation potential and spore populations of arbuscular mycorrhizal fungi. *Agriculture Ecosystems & Environment* 139 (1-2): 273-279. https://doi.org/10.1016/j.agee.2010.08.013
- Grümberg, B. C., Urcelay, C., Shroeder, M. A., Vargas-Gil, S. & Luna, C. M. (2015). The role of inoculum identity in drought stress mitigation

- by arbuscular mycorrhizal fungi in soybean. *Biology and Fertility of Soils* 51 (1): 1-10. https://doi.org/10.1007/s00374-014-0942-7
- Hanin, M., Ebel, C., Ngom, M., Laplaze, L. & Masmoudi, K. (2016). New insights on plant salt tolerance mechanisms and their potential use for breeding. *Frontiers in Plant Science* 7. https://doi.org/10.3389/fpls.2016.01787
- Harrison, M. J., Dewbre, G. R. & Liu, J. (2002). A Phosphate Transporter from *Medicago truncatula* Involved in the Acquisition of Phosphate Released by Arbuscular Mycorrhizal Fungi. *The Plant Cell* 14 (10): 2413-2429. https://doi.org/10.1105/tpc.004861
- Hart, M. M., Antunes, P. M., Chaudhary, V. B. & Abbott, L. K. (2018). Fungal inoculants in the field: Is the reward greater than the risk? *Functional Ecology* 32 (1): 126-135. https://doi.org/10.1111/1365-2435.12976
- Hawkins, H., Cargill, R. I., Van Nuland, M. E., Hagen, S. C., Field, K. J., Sheldrake, M., Soudzilovskaia, N. A. & Kiers, E. T. (2023). Mycorrhizal mycelium as a global carbon pool. *Current Biology* 33 (11): R560-R573. https://doi.org/10.1016/j.cub.2023.02.027
- Heklau, H., Schindler, N., Buscot, F., Eisenhauer, N., Ferlian, O., Salcedo, L. D. P. & Bruelheide, H. (2021). Mixing tree species associated with arbuscular or ectotrophic mycorrhizae reveals dual mycorrhization and interactive effects on the fungal partners. *Ecology and Evolution* 11 (10): 5424-5440. https://doi.org/10.1002/ece3.7437
- Hijri, M. (2016). Analysis of a large dataset of mycorrhiza inoculation field trials on potato shows highly significant increases in yield. *Mycorrhiza* 26 (3): 209-214. https://doi.org/10.1007/s00572-015-0661-4
- Hildermann, I., Messmer, M., Dubois, D., Boller, T., Wiemken, A. & Mäder, P. (2010). Nutrient use efficiency and arbuscular mycorrhizal root colonisation of winter wheat cultivars in different farming systems of the DOK long-term trial. *Journal of the Science of Food and Agriculture* n/a. https://doi.org/10.1002/jsfa.4048
- Hu, D., Baskin, J. M., Baskin, C. C., Wang, Z., Zhang, S., Yang, X. & Huang, Z. (2019). Arbuscular mycorrhizal symbiosis and achene mucilage have independent functions in seedling growth of a desert shrub. *Journal of Plant Physiology* 232: 1-11. https://doi.org/10.1016/j.jplph.2018.11.010
- Hunter, M. C., Smith, R. G., Schipanski, M. E., Atwood, L. W. & Mortensen, D. A. (2017). Agriculture in 2050: Recalibrating Targets for Sustainable Intensification. *BioScience* 67 (4): 386-391. https://doi.org/10.1093/biosci/bix010
- Igiehon, N. O. & Babalola, O. O. (2017). Biofertilizers and sustainable agriculture: exploring arbuscular mycorrhizal fungi. *Applied Microbiology and Biotechnology* 101 (12): 4871-4881. https://doi.org/10.1007/s00253-017-8344-z
- Jakobsen, I., Abbott, L. K. & Robson, A. D. (1992). External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifoli*-

- um subterraneum L. New Phytologist 120 (3): 371-380. https://doi.org/10.1111/j.1469-8137.1992.tb01077.x
- Jansa, J., Erb, A., Oberholzer, H., Šmilauer, P. & Egli, S. (2014). Soil and geography are more important determinants of indigenous arbuscular mycorrhizal communities than management practices in Swiss agricultural soils. *Molecular Ecology* 23 (8): 2118-2135. https://doi.org/10.1111/mec.12706
- Jin, H., Germida, J. J. & Walley, F. L. (2013). Suppressive effects of seed-applied fungicides on arbuscular mycorrhizal fungi (AMF) differ with fungicide mode of action and AMF species. *Applied Soil Ecology* 72: 22-30. https://doi.org/10.1016/j.apsoil.2013.05.013
- Johansen, A., Jakobsen, I. & Jensen, E. S. (1993). External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifolium subterraneum* L. *New Phytologist* 124 (1): 61-68. https://doi.org/10.1111/j.1469-8137.1993.tb03797.x
- Johnson, D., Martin, F., Cairney, J. W. G. & Anderson, I. C. (2012). The importance of individuals: intraspecific diversity of mycorrhizal plants and fungi in ecosystems. *New Phytologist* 194 (3): 614-628. https://doi.org/10.1111/j.1469-8137.2012.04087.x
- Kaur, S. & Suseela, V. (2020). Unraveling arbuscular Mycorrhiza-Induced changes in plant primary and secondary metabolome. *Metabolites* 10 (8): 335. https://doi.org/10.3390/metabo10080335
- Kafle, A., Cope, K. R., Raths, R., Yakha, J. K., Subramanian, S., Bücking, H. & Garcia, K. (2019). Harnessing soil microbes to improve plant phosphate efficiency in cropping systems. *Agronomy* 9 (3): 127. https://doi.org/10.3390/agronomy9030127
- Kaiser, C., Kilburn, M. R., Clode, P. L., Fuchslueger, L., Koranda, M., Cliff, J. B., Solaiman, Z. M. & Murphy, D. V. (2014). Exploring the transfer of recent plant photosynthates to soil microbes: mycorrhizal pathway vs direct root exudation. *New Phytologist* 205 (4): 1537-1551. https://doi.org/10.1111/nph.13138
- Kenrick, P. & Strullu-Derrien, C. (2014). The origin and early evolution of roots. *Plant Physiology* 166 (2): 570-580. https://doi.org/10.1104/pp.114.244517
- Kinge, T. R., Ghosh, S., Cason, E. D. & Gryzenhout, M. (2022). Characterization of the Endophytic Mycobiome in Cowpea (*Vigna unguiculata*) from a Single Location Using Illumina Sequencing. *Agriculture* 12 (3): 333. https://doi.org/10.3390/agriculture12030333
- Kirkegaard, J. & Sarwar, M. (1998). Biofumigation potential of brassicas. Plant and Soil 201 (1): 71-89. https://doi.org/10.1023/a:1004364713152
- Kour, D., Rana, K. L., Yadav, A. N., Yadav, N., Kumar, M., Kumar, V., Vyas, P., Dhaliwal, H. S. & Saxena, A. K. (2020). Microbial biofertilizers: Bioresources and eco-friendly technologies for agricultural and environmental sustainability. *Biocatalysis and Agricultural Biotechnology* 23: 101487. https://doi.org/10.1016/j.bcab.2019.101487

- Kumar, A., Singh, R. & Adholeya, A. (2017). Biotechnological advancements in industrial production of arbuscular mycorrhizal fungi: achievements, challenges, and future prospects. In: Satyanarayana, T., Deshmukh, S. and Johri, B. (Eds.), Developments in Fungal Biology and Applied Mycology. Singapore, Springer, (pp. 413–431). https://doi.org/10.1007/978-981-10-4768-8 21
- Kuyper, T. W. & Jansa, J. (2023). Arbuscular mycorrhiza: advances and retreats in our understanding of the ecological functioning of the mother of all root symbioses. *Plant and Soil* 489 (1-2): 41-88. https://doi.org/10.1007/s11104-023-06045-z
- Laxa, M., Liebthal, M., Telman, W., Chibani, K. & Dietz, K. J. (2019). The role of the plant antioxidant system in drought tolerance. *Antioxidants* 8 (4): 94. https://doi.org/10.3390/antiox8040094
- Lehmann, J., Hansel, C. M., Kaiser, C., Kleber, M., Maher, K., Manzoni, S., Nunan, N., Reichstein, M., Schimel, J. P., Torn, M. S., Wieder, W. R. & Kögel-Knabner, I. (2020). Persistence of soil organic carbon caused by functional complexity. *Nature Geoscience* 13 (8): 529-534. https://doi.org/10.1038/s41561-020-0612-3
- Leifheit, E. F., Veresoglou, S. D., Lehmann, A., Morris, E. K. & Rillig, M. C. (2014). Multiple factors influence the role of arbuscular mycorrhizal fungi in soil aggregation—a meta-analysis. *Plant and Soil* 374 (1-2): 523-537. https://doi.org/10.1007/s11104-013-1899-2
- Lindahl, B. D., Ihrmark, K., Boberg, J., Trumbore, S. E., Högberg, P., Stenlid, J. & Finlay, R. D. (2007). Spatial separation of litter decomposition and mycorrhizal nitrogen uptake in a boreal forest. *New Phytologist* 173 (3): 611-620. https://doi.org/10.1111/j.1469-8137.2006.01936.x
- Linderman, R. G. & Davis, E. A. (2001). Comparative Response of Selected Grapevine Rootstocks and Cultivars to Inoculation with Different Mycorrhizal Fungi. *American Journal of Enology and Viticulture* 52 (1): 8-11. https://doi.org/10.5344/ajev.2001.52.1.8
- López-Ráez, J. A., Pozo, M. J. & García-Garrido, J. M. (2011). Strigolactones: a cry for help in the rhizosphere. *Botany* 89 (8): 513-522. https://doi.org/10.1139/b11-046
- López-Ráez, J. A., Shirasu, K. & Foo, E. (2017). Strigolactones in Plant Interactions with Beneficial and Detrimental Organisms: The Yin and Yang. *Trends in Plant Science* 22 (6): 527-537. https://doi.org/10.1016/j.tplants.2017.03.011
- Luginbuehl, L. H., Menard, G. N., Kurup, S., Van Erp, H., Radhakrishnan, G. V., Breakspear, A., Oldroyd, G. E. D. & Eastmond, P. J. (2017). Fatty acids in arbuscular mycorrhizal fungi are synthesized by the host plant. *Science* 356 (6343): 1175-1178. https://doi.org/10.1126/science.aan0081
- Madawala, H. M. S. P. (2021). Arbuscular mycorrhizal fungi as biofertilizers: Current trends, challenges, and future prospects. In: Rakshit, A., Meena, V.S., Parihar, M., Singh, H.B. and Singh, A.K (Eds.), Biofertil-

- izers: Advances in Bio-Inoculants. Cambridge, Woodhead Publishing (pp. 83–93). https://doi.org/10.1016/b978-0-12-821667-5.00029-4
- Martín-Rodríguez, J. A., Huertas, R., Ho-Plágaro, T., Ocampo, J. A., Turečková, V., Tarkowská, D., Ludwig-Müller, J. & García-Garrido, J. M. (2016). Gibberellin-Abscisic Acid Balances during Arbuscular Mycorrhiza Formation in Tomato. Frontiers in Plant Science 7. https://doi.org/10.3389/fpls.2016.01273
- Miransari, M. (2011). Hyperaccumulators, arbuscular mycorrhizal fungi and stress of heavy metals. *Biotechnology Advances* 29 (6): 645-653. https://doi.org/10.1016/j.biotechadv.2011.04.006
- Miransari, M., Bahrami, H., Rejali, F. & Malakouti, M. (2009). Effects of arbuscular mycorrhiza, soil sterilization, and soil compaction on wheat (*Triticum aestivum* L.) nutrients uptake. *Soil and Tillage Research* 104 (1): 48-55. https://doi.org/10.1016/j.still.2008.11.006
- Mukhongo, R. W., Tumuhairwe, J. B., Ebanyat, P., AbdelGadir, A. A. H., Thuita, M. N., & Masso, C. (2016). Production and use of arbuscular mycorrhizal fungi inoculum in Sub-Saharan Africa: Challenges and ways of improving. *International Journal of Soil Science* 11 (3): 108-122. https://doi.org/10.3923/ijss.2016.108.122
- Nadeem, S. M., Ahmad, M., Zahir, Z. A., Javaid, A. & Ashraf, M. (2014). The role of mycorrhizae and plant growth promoting rhizobacteria (PGPR) in improving crop productivity under stressful environments. *Biotechnology Advances* 32 (2): 429-448. https://doi.org/10.1016/j.biotechadv.2013.12.005
- Naik, K., Mishra, S., Srichandan, H., Singh, P. K. & Choudhary, A. (2020). Microbial formulation and growth of cereals, pulses, oilseeds and vegetable crops. *Sustainable Environment Research* 30 (1). https://doi.org/10.1186/s42834-020-00051-x
- Neumann, E. & George, E. (2010). Nutrient uptake: The arbuscular mycorrhiza fungal symbiosis as a plant nutrient acquisition strategy. In: Koltai, H., Kapulnik, Y. (Eds.), Arbuscular Mycorrhizas: Physiology and Function. Dordrecht, Springer, (pp. 137–167). https://doi.org/10.1007/978-90-481-9489-6\_7
- Nguvo, K. J. & Gao, X. (2019). Weapons hidden underneath: bio-control agents and their potentials to activate plant induced systemic resistance in controlling crop Fusarium diseases. *Journal of Plant Diseases and Protection* 126 (3): 177-190. https://doi.org/10.1007/s41348-019-00222-y
- Njeru, E. M., Avio, L., Bocci, G., Sbrana, C., Turrini, A., Bàrberi, P., Giovannetti, M. & Oehl, F. (2015). Contrasting effects of cover crops on 'hot spot' arbuscular mycorrhizal fungal communities in organic tomato. *Biology and Fertility of Soils* 51 (2): 151-166. https://doi.org/10.1007/s00374-014-0958-z
- Nosheen, S., Ajmal, I. & Song, Y. (2021). Microbes as biofertilizers, a potential approach for sustainable crop production. *Sustainability* 13 (4): 1868. https://doi.org/10.3390/su13041868

- Nziguheba, G. & Smolders, E. (2008). Inputs of trace elements in agricultural soils via phosphate fertilizers in European countries. *The Science of the Total Environment* 390 (1): 53-57. https://doi.org/10.1016/j.scitotenv.2007.09.031
- Ouledali, S., Ennajeh, M., Ferrandino, A., Khemira, H., Schubert, A. & Secchi, F. (2019). Influence of arbuscular mycorrhizal fungi inoculation on the control of stomata functioning by abscisic acid (ABA) in drought-stressed olive plants. *South African Journal of Botany* 121: 152-158. https://doi.org/10.1016/j.sajb.2018.10.024
- Paszkowski, U., Kroken, S., Roux, C. & Briggs, S. P. (2002). Rice phosphate transporters include an evolutionarily divergent gene specifically activated in arbuscular mycorrhizal symbiosis. *Proceedings of the National Academy of Sciences* 99 (20): 13324-13329. https://doi.org/10.1073/pnas.202474599
- Patanita, M., Campos, M. D., Félix, M. D. R., Carvalho, M. & Brito, I. (2020). Effect of Tillage System and Cover Crop on Maize Mycorrhization and Presence of *Magnaporthiopsis maydis*. *Biology* 9 (3): 46. https://doi.org/10.3390/biology9030046
- Pellegrino, E., Gamper, H. A., Ciccolini, V. & Ercoli, L. (2020). Forage rotations conserve diversity of arbuscular mycorrhizal fungi and soil fertility. *Frontiers in Microbiology* 10. https://doi.org/10.3389/fmicb.2019.02969
- Powell, J. R. & Rillig, M. C. (2018). Biodiversity of arbuscular mycorrhizal fungi and ecosystem function. *New Phytologist* 220 (4): 1059-1075. https://doi.org/10.1111/nph.15119
- Qin, Z., Tian, Y., Hao, W., Zhang, J., Feng, G., Christie, P. & Gai, J. (2024). Identifying the predictors of mycorrhizal response under multiple fertilization regimes. *Agriculture Ecosystems & Environment* 365: 108926. https://doi.org/10.1016/j.agee.2024.108926
- Rausch, C., Daram, P., Brunner, S., Jansa, J., Laloi, M., Leggewie, G., Amrhein, N. & Bucher, M. (2001). A phosphate transporter expressed in arbuscule-containing cells in potato. *Nature* 414 (6862): 462-465. https://doi.org/10.1038/35106601
- Redecker, D., Schüßler, A., Stockinger, H., Stürmer, S. L., Morton, J. B. & Walker, C. (2013). An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). *Mycorrhiza* 23 (7): 515-531. https://doi.org/10.1007/s00572-013-0486-y
- Rivero, J., Gamir, J., Aroca, R., Pozo, M. J. & Flors, V. (2015). Metabolic transition in mycorrhizal tomato roots. *Frontiers in Microbiology* 6. https://doi.org/10.3389/fmicb.2015.00598
- Rouphael, Y., Franken, P., Schneider, C., Schwarz, D., Giovannetti, M., Agnolucci, M., De Pascale, S., Bonini, P. & Colla, G. (2015). Arbuscular mycorrhizal fungi act as biostimulants in horticultural crops. *Scientia Horticulturae* 196: 91-108. https://doi.org/10.1016/j.scienta.2015.09.002
- Sabia, E., Claps, S., Morone, G., Bruno, A., Sepe, L. & Aleandri, R. (2015). Field inoculation of arbuscular mycorrhiza on maize (*Zea mays* L.)

- under low inputs: preliminary study on quantitative and qualitative aspects. *Italian Journal of Agronomy* 10 (1): 30-33. https://doi.org/10.4081/ija.2015.607
- Sagar, A., Rathore, P., Ramteke, P. W., Ramakrishna, W., Reddy, M. S. & Pecoraro, L. (2021). Plant growth promoting rhizobacteria, arbuscular mycorrhizal fungi and their synergistic interactions to counteract the negative effects of saline soil on agriculture: key macromolecules and mechanisms. *Microorganisms* 9 (7): 1491. https://doi.org/10.3390/microorganisms9071491
- Säle, V., Aguilera, P., Laczko, E., Mäder, P., Berner, A., Zihlmann, U., Van Der Heijden, M. G. & Oehl, F. (2015). Impact of conservation tillage and organic farming on the diversity of arbuscular mycorrhizal fungi. *Soil Biology and Biochemistry* 84: 38-52. https://doi.org/10.1016/j.soil-bio.2015.02.005
- Samantaray, A., Chattaraj, S., Mitra, D., Ganguly, A., Kumar, R., Gaur, A., Mohapatra, P. K., De Los Santos-Villalobos, S., Rani, A. & Thatoi, H. (2024). Advances in microbial based bio-inoculum for amelioration of soil health and sustainable crop production. *Current Research in Microbial Sciences* 7: 100251. https://doi.org/10.1016/j.crmicr.2024.100251
- Sanmartín, N., Sánchez-Bel, P., Pastor, V., Pastor-Fernández, J., Mateu, D., Pozo, M. J., Cerezo, M. & Flors, V. (2020). Root-to-shoot signalling in mycorrhizal tomato plants upon *Botrytis cinerea* infection. *Plant Science* 298: 110595. https://doi.org/10.1016/j.plantsci.2020.110595
- Santander, C., Aroca, R., Cartes, P., Vidal, G. & Cornejo, P. (2021). Aquaporins and cation transporters are differentially regulated by two arbuscular mycorrhizal fungi strains in lettuce cultivars growing under salinity conditions. *Plant Physiology and Biochemistry* 158: 396-409. https://doi.org/10.1016/j.plaphy.2020.11.025
- Sawers, R. J. H., Svane, S. F., Quan, C., Grønlund, M., Wozniak, B., Gebreselassie, M., González-Muñoz, E., Montes, R. a. C., Baxter, I., Goudet, J., Jakobsen, I. & Paszkowski, U. (2017). Phosphorus acquisition efficiency in arbuscular mycorrhizal maize is correlated with the abundance of root-external hyphae and the accumulation of transcripts encoding PHT1 phosphate transporters. *New Phytologist* 214 (2): 632-643. https://doi.org/10.1111/nph.14403
- Schliemann, W., Ammer, C. & Strack, D. (2008). Metabolite profiling of mycorrhizal roots of *Medicago truncatula*. *Phytochemistry* 69 (1): 112-146. https://doi.org/10.1016/j.phytochem.2007.06.032
- Shahwar, D., Mushtaq, Z., Mushtaq, H., Alqarawi, A. A., Park, Y., Alshahrani, T. S. & Faizan, S. (2023). Role of microbial inoculants as bio fertilizers for improving crop productivity: A review. *Heliyon* 9 (6): e16134. https://doi.org/10.1016/j.heliyon.2023.e16134
- Sharma, S., Prasad, R., Varma, A. & Sharma, A. K. (2017). Glycoprotein Associated with Funneliformis coronatum, Gigaspora margarita and Acaulospora scrobiculata Suppress the Plant Pathogens In vitro. Asian

- Journal of Plant Pathology 11 (4): 199-202. https://doi.org/10.3923/ajp-paj.2017.199.202
- Smith, S. E. & Smith, F. A. (2011). Roles of Arbuscular Mycorrhizas in Plant Nutrition and Growth: New Paradigms from Cellular to Ecosystem Scales. *Annual Review of Plant Biology* 62 (1): 227-250. https://doi.org/10.1146/annurev-arplant-042110-103846
- Song, Y., Chen, D., Lu, K., Sun, Z. & Zeng, R. (2015). Enhanced tomato disease resistance primed by arbuscular mycorrhizal fungus. *Frontiers in Plant Science* 6. https://doi.org/10.3389/fpls.2015.00786
- Spatafora, J. W., Chang, Y., Benny, G. L., Lazarus, K., Smith, M. E., Berbee, M. L., Bonito, G., Corradi, N., Grigoriev, I., Gryganskyi, A., James, T. Y., O'Donnell, K., Roberson, R. W., Taylor, T. N., Uehling, J., Vilgalys, R., White, M. M. & Stajich, J. E. (2016). A phylum-level phylogenetic classification of zygomycete fungi based on genome-scale data. *Mycologia* 108 (5): 1028-1046. https://doi.org/10.3852/16-042
- Stephenson, J., Crane, S. F., Levy, C. & Maslin, M. (2013). Population, development, and climate change: links and effects on human health. *The Lancet* 382 (9905): 1665-1673. https://doi.org/10.1016/s0140-6736(13)61460-9
- Thompson, J. P., Clewett, T. G. & Fiske, M. L. (2013). Field inoculation with arbuscular-mycorrhizal fungi overcomes phosphorus and zinc deficiencies of linseed (*Linum usitatissimum*) in a vertisol subject to long-fallow disorder. *Plant and Soil 371* (1-2): 117-137. https://doi.org/10.1007/s11104-013-1679-z
- Tiamtanong, S., Sinma, K., Mala, T., Rungcharoenthong, P. & Amkha, S. (2015). Effects of Mycorrhizal Fungi with Phosphate Fertilizer Applications on Phosphate Solubilizing and Soil Properties of Grapes Orchard. *Modern Applied Science* 9 (1). https://doi.org/10.5539/mas.v9n1p149
- Van Der Heijden, M. G. A., Martin, F. M., Selosse, M. & Sanders, I. R. (2015). Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* 205 (4): 1406-1423. https://doi.org/10.1111/nph.13288
- Verbruggen, E., Van Der Heijden, M. G. A., Weedon, J. T., Kowalchuk, G. A. & Röling, W. F. M. (2012). Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in agricultural soils. *Molecular Ecology* 21 (10): 2341-2353. https://doi.org/10.1111/j.1365-294x.2012.05534.x
- Vierheilig, H., Steinkellner, S., Khaosaad, T. & Garcia-Garrido, J. M. (2008). The biocontrol effect of mycorrhization on soilborne fungal pathogens and the autoregulation of the AM symbiosis: one mechanism, two effects? In *Mycorrhiza* (pp. 307–320). https://doi.org/10.1007/978-3-540-78826-3 15
- Vyas, S. C. & Vyas, S. (2000). Effect of agrochemicals on mycorrhizae. In: Mukerji, K.G., Chamola, B.P. and Singh, J (Eds.), Mycorrhizal Biology.

- Boston, Springer, (pp. 289–327). https://doi.org/10.1007/978-1-4615-4265-0 18
- Wahid, F., Sharif, M., Fahad, S., Ali, A., Adnan, M., Rafiullah, N., Saud, S., Danish, S., Ali, M. A., Ahmed, N., Arslan, H., Arslan, D., Erman, M., Sabagh, A. E., Gholizadeh, F. & Datta, R. (2022). Mycorrhiza and phosphate solubilizing bacteria: Potential bioagents for sustainable phosphorus management in agriculture. *Phyton* 91 (2): 257-278. https://doi.org/10.32604/phyton.2022.016512
- Walder, F., Niemann, H., Natarajan, M., Lehmann, M. F., Boller, T. & Wiemken, A. (2012). Mycorrhizal Networks: Common Goods of Plants Shared under Unequal Terms of Trade. *Plant Physiology* 159 (2): 789-797. https://doi.org/10.1104/pp.112.195727
- Wang, W., Shi, J., Xie, Q., Jiang, Y., Yu, N. & Wang, E. (2017). Nutrient exchange and regulation in arbuscular mycorrhizal symbiosis. *Molecular Plant* 10 (9): 1147-1158. https://doi.org/10.1016/j.molp.2017.07.012
- Wang, Y., Wang, M., Li, Y., Wu, A. & Huang, J. (2018). Effects of arbuscular mycorrhizal fungi on growth and nitrogen uptake of *Chrysanthemum morifolium* under salt stress. *PLoS ONE* 13 (4): e0196408. https://doi.org/10.1371/journal.pone.0196408
- Waters, M. T., Gutjahr, C., Bennett, T. & Nelson, D. C. (2017). Strigolactone Signaling and Evolution. *Annual Review of Plant Biology* 68 (1): 291-322. https://doi.org/10.1146/annurev-arplant-042916-040925
- Yadav, R., Ror, P., Rathore, P., Kumar, S. & Ramakrishna, W. (2021). Bacillus subtilis CP4, isolated from native soil in combination with arbuscular mycorrhizal fungi promotes biofortification, yield and metabolite production in wheat under field conditions. Journal of Applied Microbiology 131 (1): 339-359. https://doi.org/10.1111/jam.14951
- Yooyongwech, S., Phaukinsang, N., Cha-Um, S. & Supaibulwatana, K. (2013). Arbuscular mycorrhiza improved growth performance in *Macadamia tetraphylla* L. grown under water deficit stress involves soluble sugar and proline accumulation. *Plant Growth Regulation* 69 (3): 285-293. https://doi.org/10.1007/s10725-012-9771-6
- Yu, L., Zhang, W., Geng, Y., Liu, K. & Shao, X. (2022). Cooperation with arbuscular mycorrhizal fungi increases plant nutrient uptake and improves defenses against insects. *Frontiers in Ecology and Evolution* 10. https://doi.org/10.3389/fevo.2022.833389
- Zeng, L., Jian-fu, L., Jian-fu, L. & Ming-yuan, W. (2014). Effects of arbuscular mycorrhizal (AM) fungi on citrus quality under nature conditions. *Southwest China Journal of Agricultural Sciences* 27 (5): 2101-2105. https://doi.org/10.16213/j.cnki.scjas.2014.05.067
- Zhang, J., Zhao, R., Li, X. & Zhang, J. (2024). Potential of arbuscular my-corrhizal fungi for soil health: A review. *Pedosphere* 34 (2): 279-288. https://doi.org/10.1016/j.pedsph.2024.02.002

Zhang, X., Dong, W., Sun, J., Feng, F., Deng, Y., He, Z., Oldroyd, G. E. & Wang, E. (2015). The receptor kinase CERK1 has dual functions in symbiosis and immunity signalling. *The Plant Journal* 81 (2): 258-267. https://doi.org/10.1111/tpj.12723